



Contents lists available at ScienceDirect

Biochemical and Biophysical Research Communications

journal homepage: www.elsevier.com/locate/ybbrc



Isoflurane induced cognitive impairment in aged rats through hippocampal calcineurin/NFAT signaling



Cheng Ni, Zhengqian Li, Min Qian, Yang Zhou, Jun Wang, Xiangyang Guo*

Department of Anesthesiology, Peking University Third Hospital, Beijing 100191, China

ARTICLE INFO

Article history:

Received 8 January 2015

Available online 1 April 2015

Keywords:

Calcineurin

Nuclear factor of activated T-cells

Isoflurane

Cyclosporine A

Cognitive impairment

ABSTRACT

Calcineurin (CaN) over-activation constrains synaptic plasticity and memory formation. Upon CaN activation, NFAT imports into the nucleus and guides its downstream genes, which also affect neuronal and synaptic function. Aberrant CaN/NFAT signaling involves in neurotoxicity and cognitive impairment in neurological disorders such as Alzheimer's disease, but its role in postoperative cognitive dysfunction (POCD) remains uninvestigated. Inhaled anesthetic isoflurane facilitates the development of POCD, and the present study investigated the role of CaN/NFAT signaling in isoflurane induced cognitive impairment of aged rats, and the therapeutic effects of CaN inhibitor cyclosporine A (CsA). The results indicated that hippocampal CaN activity increased and peaked at 6 h after isoflurane exposure, and NFAT, especially NFATc4, imported into the nucleus following CaN activation. Furthermore, pharmacological inhibition of CaN by CsA markedly attenuated isoflurane induced aberrant CaN/NFATc4 signaling in the hippocampus, and rescued relevant spatial learning and memory impairment of aged rats. Overall, the study suggests hippocampal CaN/NFAT signaling as the upstream mechanism of isoflurane induced cognitive impairment, and provides potential therapeutic target and possible treatment methods for POCD.

© 2015 Elsevier Inc. All rights reserved.

1. Introduction

Calcineurin (CaN) is a calcium/calmodulin-dependent serine/threonine protein kinase, which is sensitive to intracellular calcium level change. CaN is required for synaptic responses [1] and function [2], and could be a regulator for neuronal connectivity. Nuclear factor of activated T-cells (NFAT) imports into nucleus following CaN mediated dephosphorylation, and it also involves in axonal outgrowth [3], neuronal response [4], and synaptic development and plasticity [5]. While CaN/NFAT signaling is important for memory formation, its over-activation may also impair learning and memory. It has been reported that Alzheimer's disease (AD) associates with aberrant hippocampal CaN/NFAT signaling [6], and A β peptides could over-activate CaN activity in neurons [7]. Meanwhile, aberrant CaN/NFAT signaling is linked to pathologies such as synaptic dysfunction, astrocyte activation and neuronal death [8,9], and CaN inhibitor has been reported to attenuate relevant pathologies and improve cognition in AD model [10].

Postoperative cognitive dysfunction (POCD) is a major clinical issue in geriatric surgical patients [11]. It is self-limiting in most patients, but in some patients, it is long-term or even permanent [12]. POCD is associated with increased disability and early mortality [13]. Commonly used inhaled anesthetic isoflurane facilitates POCD and relevant neurotoxicity [14], but unfortunately, its upstream mechanism remains elusive. Isoflurane could induce over-activation of inositol 1,4,5-trisphosphate or ryanodine (IP3R) receptors on endoplasmic reticular (ER) membrane [15], and result in calcium leakage from ER [16]. Isoflurane could also increase bax/bcl-2 ratio and activate caspase 3 [17], which in turn cleaves IP3R and results in persistent calcium leakage [18]. These ER and mitochondria dependent pathways contribute to cytosolic calcium concentration elevation. Thus, isoflurane may lead to activation of CaN/NFAT signaling, but to our knowledge, the role of CaN/NFAT signaling in POCD remains uninvestigated.

Based on above-mentioned results and the similarity between AD and POCD, we hypothesize CaN/NFAT signaling involves in the development of POCD. And the present study aims to elucidate whether isoflurane activates hippocampal CaN/NFAT signaling and the involved NFAT isoform, furthermore, to evaluate the therapeutic effects of CaN inhibitor on isoflurane induced aberrant CaN/NFAT signaling and cognitive impairment.

* Corresponding author. Fax: +86 10 82267276.

E-mail address: puthmzk@163.com (X. Guo).

2. Materials and methods

2.1. Animals

Aged male Sprague–Dawley rats, 18 month old, weighing 550–600 g were used for the experiments. Before experiments, they were maintained on a standard housing condition with food and water ad libitum for 2 weeks.

2.2. Experiment protocols

The experimental protocols were approved by the Peking University Biomedical Ethics Committee Experimental Animal Ethics Branch (Approval No. LA201412). To investigate the effects of isoflurane exposure on CaN activity and NFAT nuclear import in the hippocampus, rats were randomly divided into isoflurane or control groups and received isoflurane (Baxter, Deerfield, IL, USA) or vehicle gas. CaN expression and activity were assessed with western blot and colorimetric analysis at 0, 3, 6, 12 and 24 h after isoflurane exposure ($n = 6$). Then NFATc2, c3 and c4 nuclear imports were observed at the most apparent CaN activation time point.

To investigate the role of hippocampal CaN/NFAT signaling in isoflurane induced cognitive dysfunction, rats were randomly divided into control, cyclosporin A (CsA, CaN inhibitor [19]), isoflurane or isoflurane + CsA groups. Rats in CsA and isoflurane + CsA groups received intraperitoneal injection of CsA (Abcam, Cambridge, UK) 7 mg/kg at 30 min before isoflurane exposure, and rats in other groups received normal saline. Then CaN activity and NFAT nuclear import were assessed, and hippocampus dependent spatial learning and memory function was assessed by Morris water maze test.

2.3. Isoflurane exposure

Isoflurane exposure was performed according to our previous study [20]. Briefly, rats received isoflurane, with 4 h treatment time in an anesthetic chamber. Isoflurane concentrations were monitored from gas outlet. SaO₂, heart rate, blood pressure, and rectal temperature were also monitored. After exposure, rats received 100% oxygen until regaining consciousness. Isoflurane was well tolerated; meanwhile SaO₂, heart rate, blood pressure and rectal temperature remained within physiological range. Rats in control and CsA groups just received vehicle gas.

2.4. Western blot

Western blot was performed to determine CaN expression in the hippocampus. Briefly, hippocampus and lysis buffer were homogenized and centrifuged, and total protein concentration of the supernatant was determined using BCA assay. Electrophoresis of protein lysates was performed on SDS–PAGE, and separated proteins were transferred to membranes and nonspecific binding sites were blocked. Then, membranes were incubated in anti-CaN antibody (1: 1000; Abcam, Cambridge, UK) or anti- β -actin antibody (1:10,000; Santa Cruz Biotechnology, Inc., Santa Cruz, CA), and incubated in fluorescently labeled secondary antibody (1:10,000; LI-COR, Lincoln, NE). Immunoreactivity was visualized by scanning membranes in an Odyssey infrared imaging system (LI-COR, Lincoln, NE). For densitometric analysis, data were calculated as a ratio of ChAT/actin.

2.5. Colorimetric assay

Colorimetric assay was performed to determine CaN activity in the hippocampus. Briefly, hippocampus and lysis buffer were

homogenized and centrifuged, and protein concentration of the supernatant was determined using BCA assay. Protein lysates and calcineurin assay buffer were mixed, calcineurin substrate was added and reaction was proceeded for 10 min. Then green assay reagent was added and OD_{620nm} was read on a microplate reader (Thermo Scientific, Waltham, MA). OD_{620nm} data were converted into released phosphate amount, and calcineurin activity was calculated as a ratio of phosphate amount/reaction time.

2.6. Immunofluorescence

Immunofluorescence staining was performed to determine NFAT nuclear import in hippocampus. Briefly, hippocampus was fixed with 4% paraformaldehyde and cryoprotected with 30% sucrose. Cryosectioning was done with Leica cryostat (Leica, Deerfield, IL) at -20°C . 10- μm -thick coronal sections were collected and incubated in anti-NFATc2, c3 or c4 antibody (1:100; Abcam, Cambridge, UK), and in FITC-conjugated secondary antibody (1:200; Santa Cruz Biotechnology, Inc., Santa Cruz, CA). Then nuclei were counterstained with DAPI (1:5000; Roche, Mannheim, Germany). Images were captured with Zeiss confocal fluorescence microscopy (Zeiss, Göttingen, Germany). As CA1 region of hippocampus plays an important role in memory formation [21], this region was analyzed.

2.7. Morris water maze

Spatial learning and memory was evaluated by Morris water maze. Place navigation test was performed 24 h after isoflurane exposure, during which rats received four training trials daily for 5 days, and during each trial, rats were placed in water facing the wall of maze at one of four equally spaced start positions. The time to locate submerged platform (escape latency, defined by cut-off time of 120 s) and the swim speed were recorded. Probe test (120 s) was performed 24 h after last trial, during which the platform was removed. The target zone transitions, target quadrant dwell time and swim path were recorded.

2.8. Statistical analysis

Statistical analysis was performed with Graphpad Prism 5.0 software. Data were expressed as means \pm SD. One-way or two-way analysis of variance (ANOVA) was used to compare CaN expression and activity, and probe test results. Two-way repeated-measures ANOVA followed by post-hoc Bonferroni test was used to compare place navigation test results. Two-tailed test was employed in all comparisons. $P < 0.05$ was considered statistically significant.

3. Results

The expression and activity of CaN in the hippocampus of aged rats were examined over a 24 h period after isoflurane exposure, and the results show that CaN cleavage and the ratio of cleaved CaN to full length CaN increased significantly ($n = 6$). Specifically, it increased at 3 h after exposure, peaked at 6 h, and persisted in a relative high level at 12 and 24 h (Fig. 1A and B). Meanwhile, CaN activity increased after isoflurane exposure ($n = 6$). Similarly, it increased at 3 h after exposure, peaked at 6 h, and persisted in a relative high level at 12 and 24 h (Fig. 1C).

As CaN activation mediates NFAT nuclear import, and NFAT plays a role in synaptic plasticity and memory formation, NFAT nuclear import was observed. The observation point was selected at 6 h after isoflurane exposure based on the change of CaN activity. In the control condition, NFATc2, c3 and c4 were primarily distributed in the cytosol of pyramidal cell layer, CA1 region of hippocampus.

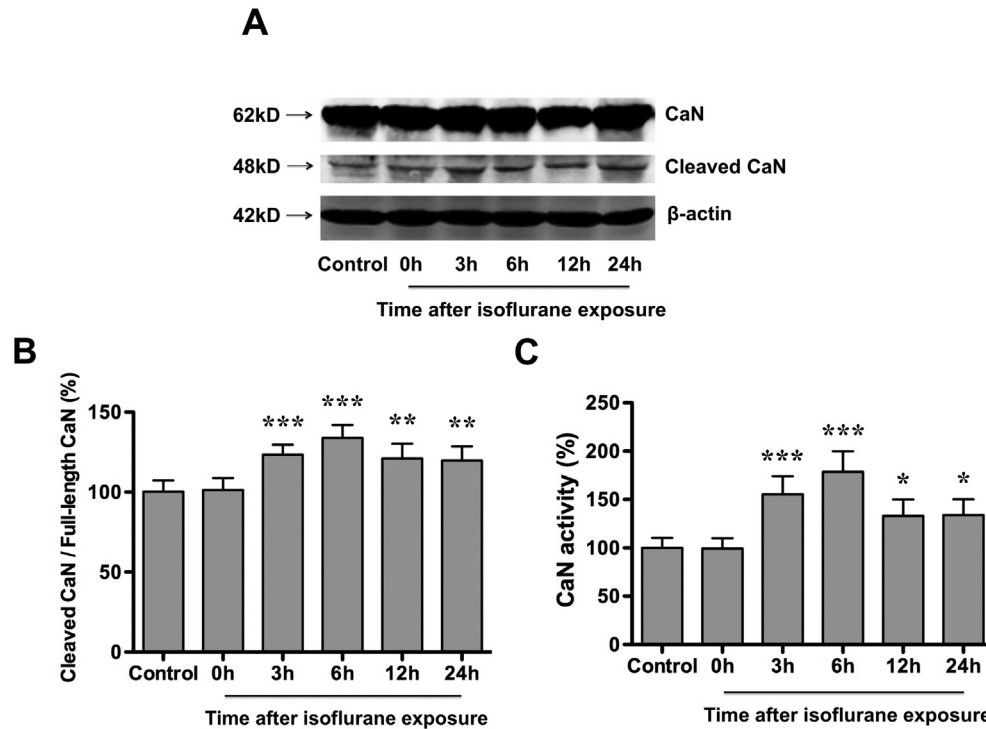


Fig. 1. Effect of 4 h isoflurane exposure on CaN cleavage and activity in the hippocampus of aged rats. (A and B) The hippocampal CaN cleavage increased significantly over time after isoflurane exposure, and peaked at 6 h (C) CaN activity also increased significantly over time after exposure, and peaked at 6 h. Data are expressed as means \pm SD ($n = 6$). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs control group.

After isoflurane exposure, NFATc4 showed apparent nuclear distribution, but NFATc2 and c3 nuclear distributions were relative few (Fig. 2). These results indicate that isoflurane exposure could activate hippocampal calcineurin/NFAT signaling, especially through NFATc4 activation.

CsA, an inhibitor of CaN, has been reported to protect against brain injury [22]. In the present study, CsA significantly decreased isoflurane induced CaN cleavage (6 h after exposure), but CsA alone did not affect CaN cleavage ($n = 6$, Fig. 3A and B). Meanwhile, CsA significantly decreased isoflurane induced CaN activation (6 h after exposure), and CsA alone also did not affect CaN activity ($n = 6$, Fig. 3C).

As NFATc4 showed apparent nuclear distribution after isoflurane exposure, the effect of CsA on NFATc4 nuclear import was observed. The results indicate that CsA attenuated isoflurane induced NFATc4 nuclear import, but CsA alone did not affect NFATc4 distribution, which remained primarily in the cytosol of pyramidal cell layer, CA1 region of hippocampus (Fig. 3D).

Finally, the effect of CsA on isoflurane induced cognitive impairment of aged rats was observed. Both group factor (treatment) and repeated factor (time) significantly affected escape latency ($P < 0.05$ and $P < 0.001$), and no interaction was found. The Bonferroni test shows that CsA significantly decreased isoflurane induced escape latency prolongation on the 4th and 5th days after exposure, but CsA alone did not affect escape latency ($n = 12$, Fig. 4A). There was no significant difference in the swim speed among four groups (Fig. 4B). In the probe test, CsA significantly increased both isoflurane induced target zone transitions and percentage of target quadrant dwell time reduction, but CsA alone affected neither target zone transitions nor percentage of target quadrant dwell time (Fig. 4C and D). Fig. 4E showed representative swim path of 4 groups in probe test. Taken together, these results show that CsA could rescue isoflurane induced cognitive impairment in aged rats through inhibiting aberrant hippocampal CaN/NFAT signaling.

4. Discussion

CaN is enriched in brain and could be activated by increased cytosolic calcium level of neurological disorders. Neuronal CaN has been reported to involve in synapse loss and dysfunction, neuronal vulnerability, neuroinflammation and neurodegeneration [23]. CaN constrains long-term potentiation (LTP) in the hippocampus, and over-expression of CaN inhibited transition from short-term to long-term memory [24]. CaN also blocks experience-dependent plasticity, a fundamental mechanism for learning and memory [25]. LTP is increased by genetic [26] or pharmacological CaN inhibition [27], on the contrary, long-term depression is blocked by CaN inhibition [28], which means CaN has dual effect in excitatory and inhibitory synapses. The present results show that isoflurane induced hippocampal CaN activation as well as memory impairment, which mean that CaN over-activation may play a role in the development of POCD.

CaN could dephosphorylate Ca^{2+} sensor/translocation domain of NFATc1–c4, resulting in NFAT nuclear import [29], and regulate synaptic function through this effect [30]. Basal activity of NFAT sits at intermediate level, and its reduction and elevation both affect synaptic function. In the hippocampus, synaptic activity regulates NFAT nuclear import, while NFAT and its downstream genes, such as IP3R1, also affect synaptic connections [31]. NFAT regulates pre-synaptic development and constrains long-term plasticity by dampening neuronal excitability [5]. NFAT over-expressing animals have more closed stable microtubule loops, and increasing NFAT activity in relevant neuronal circuits inhibits long-term behavioral adaptation [5].

During mild cognitive impairment, nuclear NFATc2 distribution increased in hippocampus, while nuclear NFATc4 distribution increased with intermediate to severe AD, and nuclear NFATc1 remained unchanged. In cerebellum, nuclear NFAT distributions

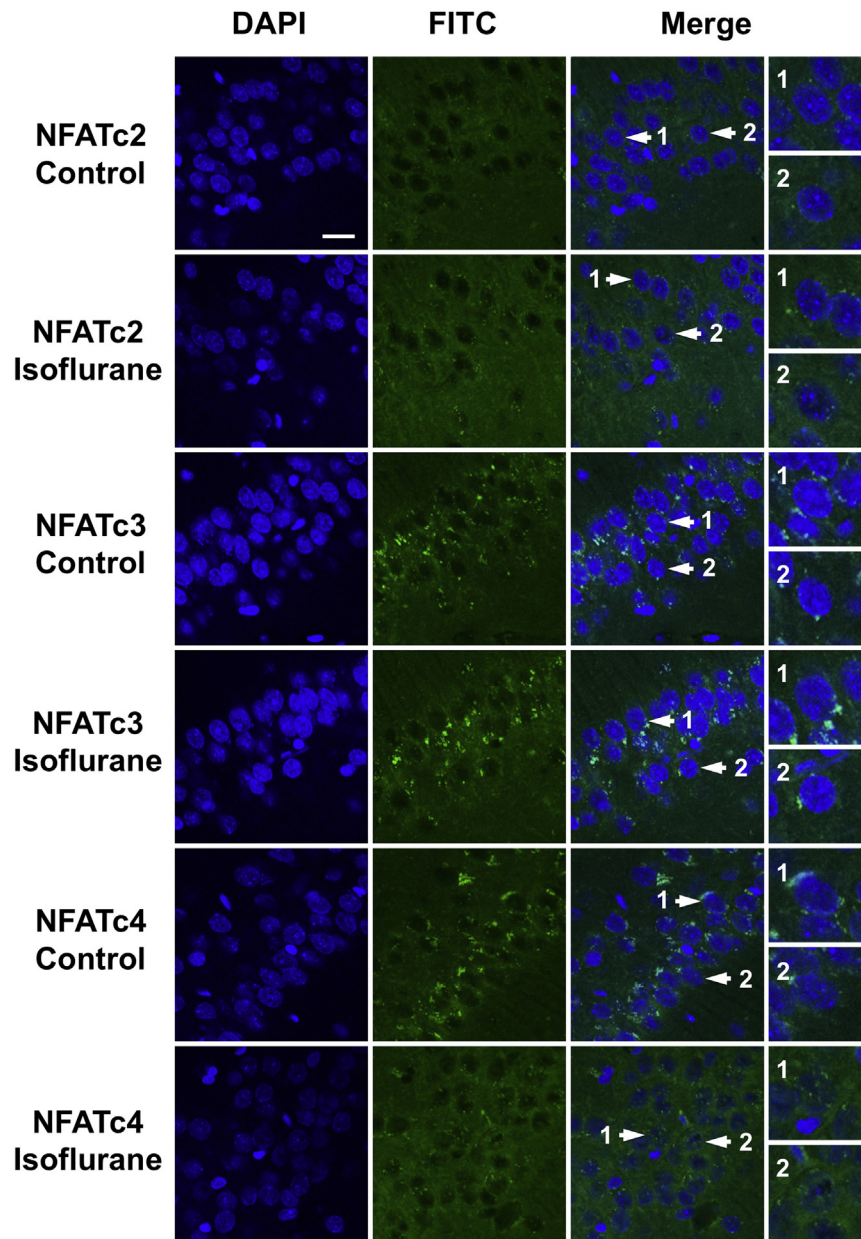


Fig. 2. Effect of 4 h isoflurane exposure on NFAT nuclear import in the hippocampus of aged rats. Confocal immunofluorescence images show double labeling for NFATc2–4 (FITC, green) and DAPI (blue) counter stain. In the control condition, NFATc2, c3 and c4 were primarily distributed in the cytosol of pyramidal cell layer, CA1 region of hippocampus. 6 h after isoflurane exposure, NFATc4 was distributed in both nucleus and cytosol, but NFATc2 and c3 nuclear distributions were relative few. In each panel, arrows point to the regions that present typical NFAT distribution, which are provided as high magnification images in the corresponding right panels. Magnification 400 \times , scale bar 20 μ m. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

were similar in different cognitive status, suggesting that NFAT involvement is restricted to vulnerable tissue [32]. In hippocampal neurons, elevations of calcium concentration rapidly induced NFATc3 nuclear import, and NFATc4 exhibited nuclear import only after prolonged (1–3 h) depolarization, which requiring coincident suppression of GSK3 β [33], and isoflurane has been reported to inhibit GSK3 β activation during brain injury [34]. We observed the nuclear imports of NFATc2, c3 and c4 in the hippocampus of aged rats, and found that 4 h isoflurane primarily induced NFATc4 nuclear import. As each NFAT isoform regulates unique transcriptional program, the results indicate that NFATc4 and its downstream genes involve in isoflurane induced synaptic dysfunction and development of POCD.

CsA is a specific CaN inhibitor. It could bind to cytosolic protein cyclophilin, and the complex of cyclosporin and cyclophilin inhibits calcineurin activity, then in step, prevents the dephosphorylation of NFAT and its nuclear import [35]. CsA crosses the blood–brain barrier (BBB) only after brain damage, such as brain ischemia [36]. We have found that isoflurane induced hippocampal BBB ultrastructure morphological damage and tight junction proteins occludin decrease, which resulted in BBB disruption and permeability increase [37]. Thus, it is possible that CsA inhibits isoflurane induced synaptic dysfunction by entering hippocampus through isoflurane induced BBB opening. The present results show that CsA attenuated isoflurane induced CaN activation and NFATc4 nuclear import, which means intraperitoneal CsA could be an effective inhibitor of aberrant

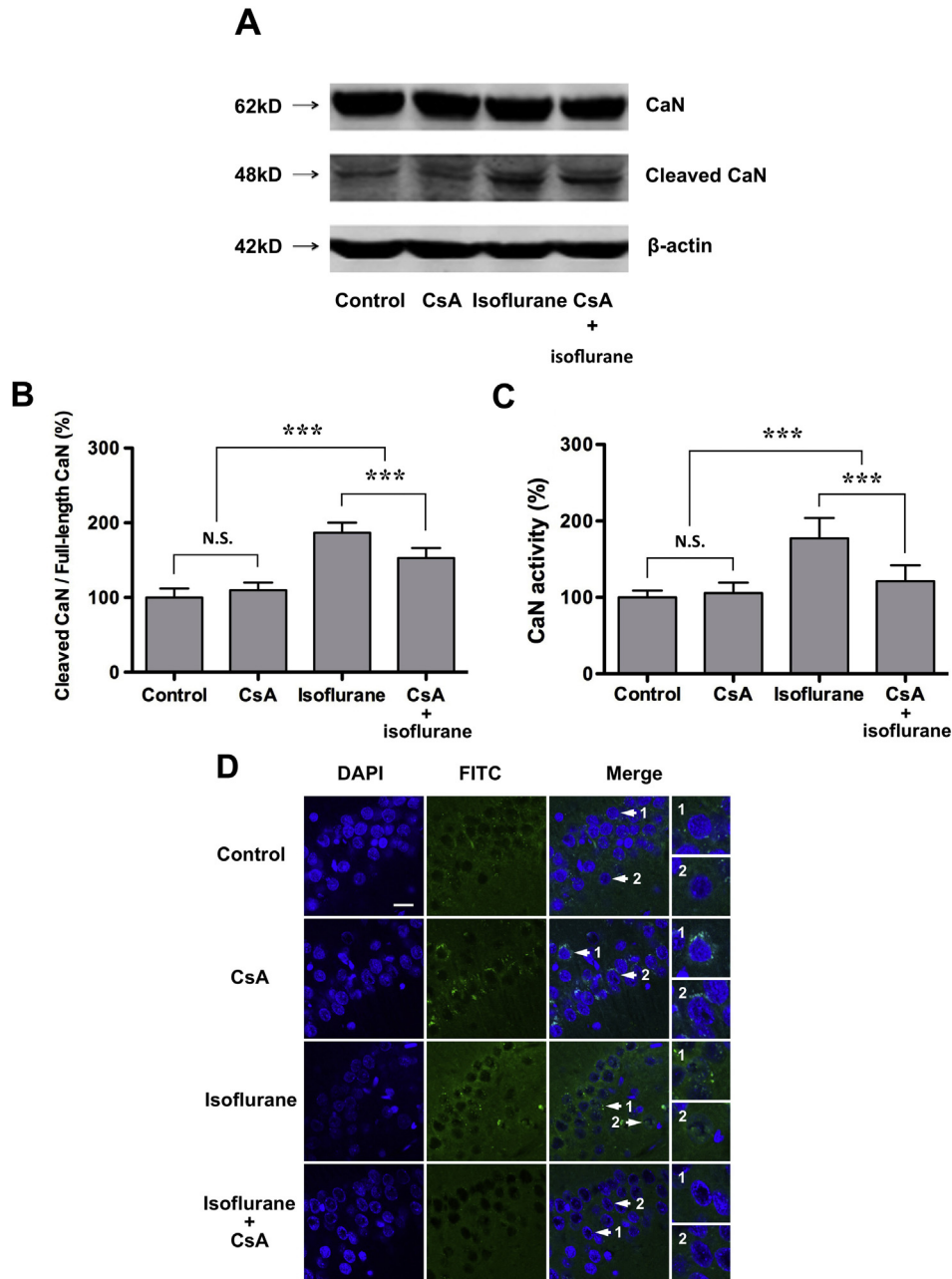


Fig. 3. CsA attenuated isoflurane exposure induced aberrant CaN/NFAT signaling in the hippocampus of aged rats. CsA 7 mg/kg was injected intraperitoneally 30 min before exposure. (A and B) The hippocampal CaN cleavage increased significantly 6 h after isoflurane exposure. CsA attenuated isoflurane induced CaN cleavage, but CsA alone did not affect CaN cleavage. (C) CaN activity increased significantly after isoflurane exposure. CsA attenuated isoflurane induced CaN activation, but CsA alone did not affect CaN activity. (D) NFATc4 nuclear distribution increased after isoflurane exposure in the pyramidal cell layer, CA1 region of hippocampus. CsA attenuated isoflurane induced NFATc4 nuclear import, but CsA alone did not affect NFATc4 distribution. In each panel, arrows point to the regions that present typical NFAT distribution, which are provided as high magnification images in the corresponding right panels. Magnification 400 \times , scale bar 20 μ m. Data are expressed as means \pm SD (n = 6). ***p < 0.001.

CaN/NFAT signaling after isoflurane exposure. Meanwhile, CsA alone had no effect on spatial memory of rats, indicating that the effect of CsA is dependent on isoflurane induced BBB opening.

Isoflurane induces cognitive impairment in aged rats [36,38,39], and the present results show that CsA rescued isoflurane induced cognitive impairment. Considering the role of CaN/NFAT signaling in synaptic plasticity and memory formation, the results illustrate that CaN/NFAT signaling is an important upstream mechanism and potential therapeutic target for isoflurane induced cognitive impairment. APP transgenic mice display increased CaN activity in brain coincident with plaque formation and cognitive impairment,

and could be treated by CaN inhibitor [40]. A β leads to aberrant CaN/NFAT activity in both neurons and astrocytes during the progression of AD [6]. Aberrant CN/NFAT also transcriptionally regulates β -site APPcleaving enzyme 1 (BACE-1), then BACE-1 promotes conversion of APP to A β , leading to the loss of neuronal function and viability [41]. We have found that isoflurane increased A β and related neurotoxicity in the hippocampus of aged rodents [14,20]. Thus, isoflurane may induce a vicious cycle of aberrant CaN/NFAT and A β increase, resulting in synaptic dysfunction and cognitive impairment, and CaN inhibitor could be an effective treatment to prevent this vicious cycle and isoflurane induced neurotoxicity.

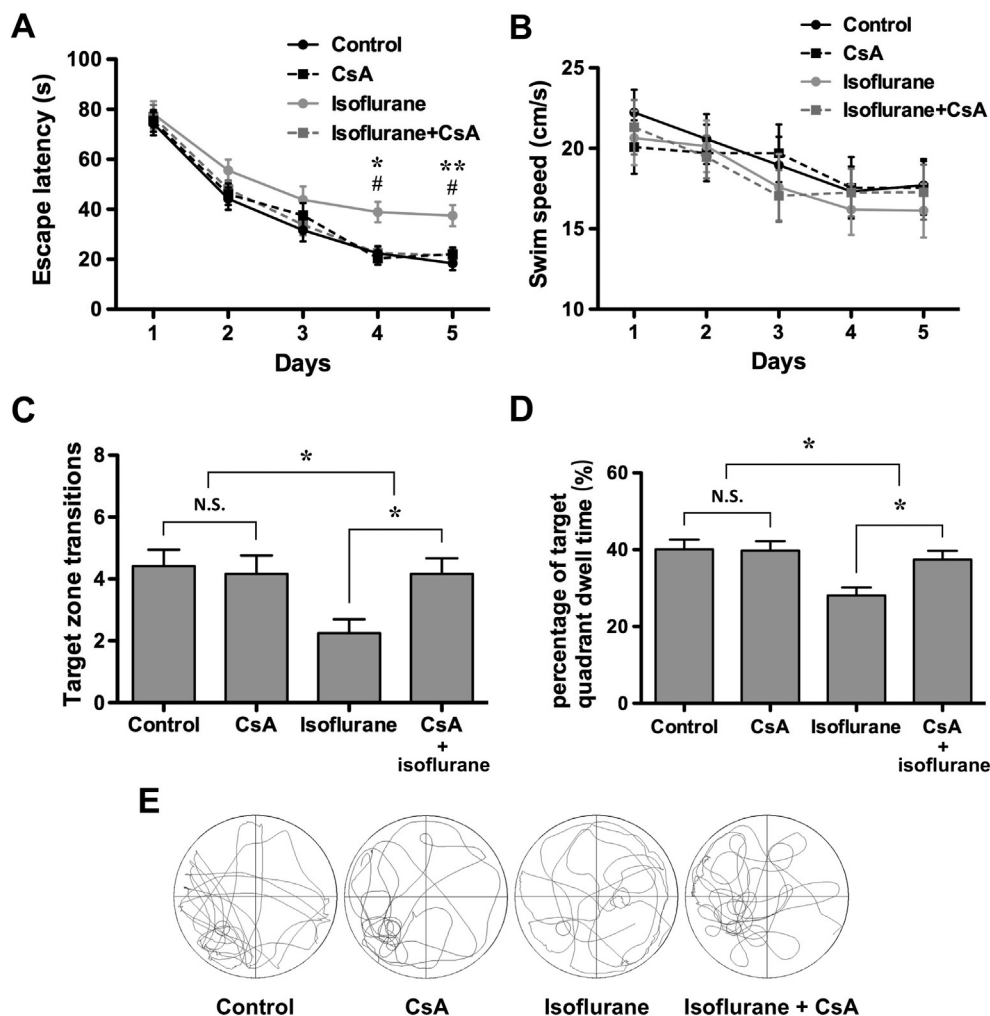


Fig. 4. CsA attenuated isoflurane exposure induced spatial memory impairment of aged rats. (A) Isoflurane increased escape latency as compared with control condition. CsA (7 mg/kg) attenuated isoflurane induced escape latency prolongation, but CsA alone did not affect escape latency (* $p < 0.05$, ** $p < 0.01$ isoflurane group vs control group, # $p < 0.05$ isoflurane + CsA group vs isoflurane group). (B) Isoflurane and CsA did not affect swim speed. (C and D) Isoflurane decreased both target zone transition and percent of target quadrant dwell time as compared with control condition. CsA attenuated isoflurane induced target zone transition and percent of target quadrant dwell time decrease, but CsA alone did not affect them (* $p < 0.05$). (E) Representative swim path of 4 groups in probe test. Data are expressed as means \pm SEM ($n = 12$).

In conclusion, the present study indicates that isoflurane induces CaN activation and NFAT, especially NFATc4, nuclear import in the hippocampus. Furthermore, CsA attenuates isoflurane induced aberrant CaN/NFAT signaling, and rescues spatial learning and memory impairment of aged rats. These results reveal an important role of hippocampal CaN/NFAT signaling in inhaled anesthetic isoflurane induced synaptic dysfunction and cognitive impairment, and provide potential therapeutic target and effective treatment methods for POCD.

Conflict of interest

We declare that we have no conflict of interest.

Acknowledgments

The present study was supported by the National Natural Science Foundation Of China (No. 81400869), and Scientific Research Foundation for Returned Scholars, Peking University Third Hospital.

Transparency document

Transparency document related to this article can be found online at <http://dx.doi.org/10.1016/j.bbrc.2015.03.083>.

References

- [1] W. Morishita, H. Marie, R.C. Malenka, Distinct triggering and expression mechanisms underlie LTD of AMPA and NMDA synaptic responses, *Nat. Neurosci.* 8 (2005) 1043–1050.
- [2] A. Shalizi, et al., A calcium-regulated MEF2 sumoylation switch controls postsynaptic differentiation, *Science* 311 (2006) 1012–1017.
- [3] I.A. Graef, et al., Neurotrophins and netrins require calcineurin/NFAT signaling to stimulate outgrowth of embryonic axons, *Cell* 113 (2003) 657–670.
- [4] R.D. Groth, P.G. Mermelstein, Brain-derived neurotrophic factor activation of NFAT (nuclear factor of activated T-cells)-dependent transcription: a role for the transcription factor NFATc4 in neurotrophin-mediated gene expression, *J. Neurosci.* 23 (2003) 8125–8134 the official Journal of Society for Neuroscience.
- [5] A. Freeman, A. Franciscovich, M. Bowers, D.J. Sandstrom, S. Sanyal, NFAT regulates pre-synaptic development and activity-dependent plasticity in *Drosophila*, *Mol. Cell. Neurosci.* 46 (2011) 535–547.
- [6] H.M. Abdul, J.L. Furman, M.A. Sama, D.M. Mathis, C.M. Norris, NFATs and Alzheimer's disease, *Mol. Cell. Pharmacol.* 2 (2010) 7–14.

- [7] L.C. Reese, W. Zhang, K.T. Dineley, R. Kayed, G. Tagliatela, Selective induction of calcineurin activity and signaling by oligomeric amyloid beta, *Aging Cell* 7 (2008) 824–835.
- [8] C.M. Norris, et al., Calcineurin triggers reactive/inflammatory processes in astrocytes and is upregulated in aging and Alzheimer's models, *J. Neurosci.* 25 (2005) 4649–4658 the official Journal of Society for Neuroscience.
- [9] H.G. Wang, et al., Ca²⁺-induced apoptosis through calcineurin dephosphorylation of BAD, *Science* 284 (1999) 339–343.
- [10] K.T. Dineley, D. Hogan, W.R. Zhang, G. Tagliatela, Acute inhibition of calcineurin restores associative learning and memory in Tg2576 APP transgenic mice, *Neurobiol. Learn. Mem.* 88 (2007) 217–224.
- [11] K.A. Hartholt, T.J. van der Cammen, M. Klimek, Postoperative cognitive dysfunction in geriatric patients, *Z. fur Gerontol. Geriatr.* 45 (2012) 411–416.
- [12] A.Y. Bekker, E.J. Weeks, Cognitive function after anaesthesia in the elderly, *Best Pract. Res. Clin. Anaesthesiol.* 17 (2003) 259–272.
- [13] J. Steinmetz, et al., Long-term consequences of postoperative cognitive dysfunction, *Anesthesiology* 110 (2009) 548–555.
- [14] Z. Xie, et al., The common inhalation anesthetic isoflurane induces caspase activation and increases amyloid beta-protein level in vivo, *Ann. Neurol.* 64 (2008) 618–627.
- [15] H. Yang, et al., Inhalational anesthetics induce cell damage by disruption of intracellular calcium homeostasis with different potencies, *Anesthesiology* 109 (2008) 243–250.
- [16] D. Lindholm, H. Wootz, L. Korhonen, ER stress and neurodegenerative diseases, *Cell Death Differ.* 13 (2006) 385–392.
- [17] Y. Zhang, et al., The mitochondrial pathway of anesthetic isoflurane-induced apoptosis, *J. Biol. Chem.* 285 (2010) 4025–4037.
- [18] Z. Assefa, et al., Caspase-3-induced truncation of type 1 inositol trisphosphate receptor accelerates apoptotic cell death and induces inositol trisphosphate-independent calcium release during apoptosis, *J. Biol. Chem.* 279 (2004) 43227–43236.
- [19] X. Wu, et al., Opposing roles for calcineurin and ATF3 in squamous skin cancer, *Nature* 465 (2010) 368–372.
- [20] C. Ni, et al., Melatonin premedication attenuates isoflurane anesthesia-induced beta-amyloid generation and cholinergic dysfunction in the hippocampus of aged rats, *Int. J. Neurosci.* 123 (2013) 213–220.
- [21] J.R. Whitlock, A.J. Heynen, M.G. Shuler, M.F. Bear, Learning induces long-term potentiation in the hippocampus, *Science* 313 (2006) 1093–1097.
- [22] M.M. Osman, et al., Cyclosporine-A as a neuroprotective agent against stroke: its translation from laboratory research to clinical application, *Neuropeptides* 45 (2011) 359–368.
- [23] L.C. Reese, G. Tagliatela, A role for calcineurin in Alzheimer's disease, *Curr. Neuropharmacol.* 9 (2011) 685–692.
- [24] I.M. Mansuy, M. Mayford, B. Jacob, E.R. Kandel, M.E. Bach, Restricted and regulated overexpression reveals calcineurin as a key component in the transition from short-term to long-term memory, *Cell* 92 (1998) 39–49.
- [25] Y. Yang, et al., Reversible blockade of experience-dependent plasticity by calcineurin in mouse visual cortex, *Nat. Neurosci.* 8 (2005) 791–796.
- [26] G. Malleret, et al., Inducible and reversible enhancement of learning, memory, and long-term potentiation by genetic inhibition of calcineurin, *Cell* 104 (2001) 675–686.
- [27] J.H. Wang, P.T. Kelly, The balance between postsynaptic Ca²⁺-dependent protein kinase and phosphatase activities controlling synaptic strength, *Learn. Mem.* 3 (1996) 170–181.
- [28] Y.M. Lu, I.M. Mansuy, E.R. Kandel, J. Roder, Calcineurin-mediated LTD of GABAergic inhibition underlies the increased excitability of CA1 neurons associated with LTP, *Neuron* 26 (2000) 197–205.
- [29] H. Okamura, et al., A conserved docking motif for CK1 binding controls the nuclear localization of NFAT1, *Mol. Cell. Biol.* 24 (2004) 4184–4195.
- [30] N. Schwartz, A. Schohl, E.S. Ruthazer, Neural activity regulates synaptic properties and dendritic structure in vivo through calcineurin/NFAT signaling, *Neuron* 62 (2009) 655–669.
- [31] I.A. Graef, et al., L-type calcium channels and GSK-3 regulate the activity of NF-ATc4 in hippocampal neurons, *Nature* 401 (1999) 703–708.
- [32] H.M. Abdul, et al., Cognitive decline in Alzheimer's disease is associated with selective changes in calcineurin/NFAT signaling, *J. Neurosci.* 29 (2009) 12957–12969 the official Journal of Society for Neuroscience.
- [33] J.D. Ulrich, et al., Distinct activation properties of the nuclear factor of activated T-cells (NFAT) isoforms NFATc3 and NFATc4 in neurons, *J. Biol. Chem.* 287 (2012) 37594–37609.
- [34] D. Lin, G. Li, Z. Zuo, Volatile anesthetic post-treatment induces protection via inhibition of glycogen synthase kinase 3beta in human neuron-like cells, *Neuroscience* 179 (2011) 73–79.
- [35] D. Tedesco, L. Haragsim, Cyclosporine: a review, *J. Transplant.* 2012 (2012) 230386.
- [36] H. Uchino, et al., Amelioration by cyclosporin A of brain damage in transient forebrain ischemia in the rat, *Brain Res.* 812 (1998) 216–226.
- [37] Y. Cao, et al., Isoflurane anesthesia results in reversible ultrastructure and occludin tight junction protein expression changes in hippocampal blood-brain barrier in aged rats, *Neurosci. Lett.* 587C (2014) 51–56.
- [38] Z.Q. Li, et al., Activation of the canonical nuclear factor-kappaB pathway is involved in isoflurane-induced hippocampal interleukin-1beta elevation and the resultant cognitive deficits in aged rats, *Biochem. Biophys. Res. Commun.* 438 (2013) 628–634.
- [39] L. Cao, L. Li, D. Lin, Z. Zuo, Isoflurane induces learning impairment that is mediated by interleukin 1beta in rodents, *PloS one* 7 (2012) e51431.
- [40] G. Tagliatela, D. Hogan, W.R. Zhang, K.T. Dineley, Intermediate- and long-term recognition memory deficits in Tg2576 mice are reversed with acute calcineurin inhibition, *Behav. Brain Res.* 200 (2009) 95–99.
- [41] H. Fukumoto, B.S. Cheung, B.T. Hyman, M.C. Irizarry, Beta-secretase protein and activity are increased in the neocortex in Alzheimer disease, *Arch. Neurol.* 59 (2002) 1381–1389.